

SUPPLEMENTAL TEXT

SUPPLEMENTAL FIGURE LEGENDS

Figure S1. Mapping the transcription start site and identifying the region footprinted by the PhoP-P protein in PhoP-activated promoters. (A) S1 nuclease protection assay of the PhoP-activated *mig-14*, *pagK*, *yrbL*, *yobG*, *pagP*, *pipD*, *ybjX*, *slyB*, *pcgL*, *pagD*, *virK*, *ompX*, *mgtC* and *pgtE* promoters carried out with mRNAs harvested from wild-type (14028s) and *phoP* (MS7953s) *Salmonella* strains following growth in N-minimal medium with 10 mM (H) or 10 μ M (L) Mg^{2+} concentrations. The arrow indicates the transcription start site. (B) DNase I footprinting analysis of promoter regions protected by the phosphorylated form of the PhoP protein (red lines) at the *mig-14*, *pagK*, *yrbL*, *yobG*, *pagP*, *pipD*, *ybjX*, *slyB*, *pcgL*, *pagD*, *virK*, *ompX*, *mgtC*, *rstA* and *pgtE* promoters. DNase I footprinting analysis has been previously reported for *mgtA* [2], *pagC* [2,3], *ugtL* [4], *phoP* [5] and *pmrD* [6].

Figure S2. DNA sequences of PhoP-activated promoters. DNA sequences corresponding to the 20 analyzed promoters. PhoP boxes are underlined and labeled by their corresponding identifier in Table S1. The central nucleotide of the PhoP box is highlighted (red color). The PhoP boxes in the reverse orientation are also presented by their reverse complementary sequence, which is the sequence format used to build the

PhoP submotifs. The RNAP -35 and -10 regions (blue color) and transcription start site (green color) are also indicated. These promoters are divided into five classes based on their shared architectures (see Fig. 4).

Figure S3. Engineered promoter constructs. Derivatives of the PhoP regulated *phoP*, *rstA*, *mgtC*, *pdgL*, *virK*, *yjbX* and *pagK* promoters. The PhoP boxes are indicated in lowercase, the RNAP -10 and -35 regions are underlined, and the transcription start site is highlighted in bold.

Figure S4. The α -CTD subunit of RNA polymerase is required to promote transcription of the *rstA* and *pagC* genes *in vitro*. Single-round *in vitro* transcription assays with linear templates corresponding to the *rstA* (A) and *pagC* (B) promoters, wild-type or mutant $\alpha \Delta 235$ ($\alpha \Delta$ -CTD) RNA polymerase, and increasing amounts of phosphorylated *Salmonella* PhoP-His-6 protein.

Figure S5. Models of interactions between a transcription factor and RNAP. (A) The CRP box, like the PhoP box, is functional when located at integral turns of the DNA helix with respect to the RNAP -10 hexamer, allowing upstream and/or downstream contacts with the α -CTD of RNAP [7]. (B) The CRP box is not functional when its face rotates 180° at half-integral turns of that hexamer [7], whereas (C) the PhoP box is

functional at that face depending on its orientation and location with respect to the RNAP -10 hexamer. **(D)** The PhoP protein is predicted to contact the σ subunit of RNAP when the PhoP box completely overlaps the RNAP -35 hexamer. **(E)** Two-fold symmetry, and inverted repeat binding site of the CRP protein [7,9,10,11]. **(F)** Two-fold symmetry regulatory domain, and direct repeat binding site of the PhoP protein [12]. Head-to-tail DNA-binding domain (e.g., PhoB protein [8]) of the PhoP protein [12].

Figure S6. Classification of ancestral and horizontally acquired PhoP-activated genes. Hierarchical clustering (Statistical Toolbox, Matlab 2007b, complete linkage method and correlation similarity were applied) of the Conservation Scores (CS) of proteins encoded by 20 PhoP-activated genes in *Salmonella*. CS is calculated as in [13] ($e\text{-value} < 1\text{E-}5$, expected number of false positives in a reciprocal BLAST search). CS values (represented by colors) can range from 55% to 100% when the closest homolog exhibits that percentage of amino acid identity. (Note that the phylogenetic relationships shown at the top of the figures do not represent phylogenetic distances.) **(A)** Group of genes exhibiting the highest CSs along the evaluated bacterial genomes. **(B)** Group of genes exhibiting the lowest CSs along the evaluated bacterial genomes. **(C)** Scatter plot combining the rank order derived from **(A)** with the CG-content (Bioinformatics Toolbox, Matlab 2007b) of the corresponding proteins in *Salmonella* ($F\text{-statistic } p\text{-value} < 4.94\text{e-}05$). Colors correspond to promoter architectures.

SUPPLEMENTAL TABLES

Table S1. Promoter dataset corresponding to 20 PhoP regulated promoters. (“Arch.” follows the promoter description of Fig. 1; “PhoP box ID” designs the PhoP box in Fig. S1; “#” accounts for the number of PhoP box(es); “O” corresponds to the PhoP box orientation; “Function box” states for the role of the PhoP boxes, where “A” and “R” correspond to activation and repression; and “Distances” describes the distances between the PhoP boxes and the RNAP binding site;).

Gene Name	Function	Arch.	ID	Sequence_PhoP-box	#	O	Function PhoP box	Distances		
								boxes	-10	+1
<i>yrbL</i>	Putative cytoplasmic protein	I	s22	TCGTTTAGGTTTGTGTTAA	1	1	A		11	27
<i>phoP</i>	Response regulator	I	s8	TGGTTTATTAACGTGTTAT	1	1	A		12	33
<i>mgtA</i>	Mg2+ transporter	I	s33	TGGTTTATCGTTGGTTTAA	1	1	A		12	33
<i>slyB</i>	Outer membrane lipoprotein	I	s12	TCGTTTAAGATTGGTTAAT	1	1	A		12	32
<i>pmrD</i>	Connector of two-component systems	I	s10	CTATTGCCGTTTGTGTTAT	1	1	A		12	31
<i>rstA</i>	Response regulator	II	s11	TCGTTTAGAAAAGATTTAT	1	1	A		23	42
<i>ompX</i>	Outer membrane protein	II	s2	CGGTTGAGGGTTCGTTGAA	1	1	A		21	42
<i>pcgL</i>	D-ala-D-ala dipeptidase	III	s27	ATTTTAACCATCTGTTTAA	2	1	A			30
<i>yobG</i>	Putative inner membrane protein	III	s62	GAGTTTATATTTGCTTAT		0	R	44		90
			s21	ACAGTTACTCCTGGTTTAA	2	1	A		12	31
			s70	GTTTTAGGAATGATTCAC		0	R	15		62
<i>pagP</i>		III	s6	CTGTTTATAGTTTGTTAAG	2	1	A		23	43
<i>pagD</i>		III	s73	TTTGTGAAAGCTTATTAAG		1	R	67		126
			s74	TGGTTAACTCTTCGTTGAA	2	1	A		22	42
			s5	GTGTTTAGAGAGAATTAC		1	R	86		144
<i>virK</i>	Putative transcription activator	IV	s17	TCGTTGCCTTTACGTTTAA	2	1	A		12	32
<i>mig-14</i>		IV	s15	CCATTGATAAACTGTTTAA		1	A	26		74
			s61	ACATTTTATTTGGTTAAG	2	1	A		14	33
			s38	ATGTTTAGCTTGATTTAA		0	A	8		57
<i>ybjX</i>		IV	s20	GTATTGACGATTGGTTAAT	2	1	A		12	31
			s18	TTGTTTAGATACGGTTTAC		0	A	30		77
<i>ugtL</i>	Inner membrane protein that modifies the LPS	V	s14	CGGTTGAGCAACTATTAC	2	0	A			51
			s28	AATAATACTTTAGTTTAA		0	R	-24		2

Gene Name	Function	Arch.	ID	Sequence_Phosphatase-box	#	O	Function Phosphatase box	Distances		
								boxes	-10	+1
<i>pagK</i>	Outer membrane protease	V	s26	CCATTTATAAAATATTTAA	2	0	A	-33	37	55
<i>pgtE</i>		V	s65	ACGTTTAATATCTATAGTA		1	R			-2
		V	s64	ATTTTTACCTTATATTGAA	2	0	A	35	49	68
		V	s7	ATGATTATAGATTGCTTAT		1	R			119
<i>mgtC</i>	Inner membrane virulence protein that aids growth in low Mg ²⁺	V	s35	CTGTTTAAGTTTGTGTTGAT	2	0	A	-368	48	67
		V	s34	ATGTTTAAACACGCTTTAT		1	R			-301
		V	s36	ATGTTTCCTTATATTTTAA		1	R	-226		-159
<i>pipD</i>	Outer membrane protein	V	s9	TTATTGAGGTTGTATTGAT	2	0	A	-131	57	76
		V	s66	CCGTTACGCTGCTGGGTAT		1	R			-55
<i>pagC</i>		V	s4	GTGTTTAGAGAGAATTTAC	2	0	A	-594	47	67
		V	s3	TTATTTACGGTGTGTTTAA		0	R			-527

Table S2. Characterization of the RNAP binding sites corresponding to 20 PhoP regulated promoters (See Methods).

Gene Name	Score -10	Score -35	Distance -35_-10	Score RNAP	Extended Element
<i>yrbL</i>	0.67	0.33	18	0.62	TGTC
<i>phoP</i>	0.83	0.67	17	0.79	
<i>mgtA</i>	0.83	0.33	17	0.74	
<i>slyB</i>	0.83	0.50	14	0.63	TGAT
<i>pmrD</i>	0.50	0.33	15	0.54	TGTG
<i>rstA</i>	0.67	0.50	17	0.75	
<i>ompX</i>	0.83	0.83	19	0.79	
<i>ybjX</i>	0.67	0.33	18	0.61	GTGG
<i>pdgL</i>	0.50	0.33	20	0.54	
<i>yobG</i>	0.50	0.33	14	0.49	ATGA
<i>pagP</i>	0.83	0.33	16	0.69	TTGT
<i>pagD</i>	0.83	0.33	17	0.72	
<i>virK</i>	0.50	0.67	16	0.70	
<i>mig-14</i>	0.67	0.50	16	0.68	
<i>ugtL</i>	0.67	0.50	15	0.66	
<i>pagK</i>	0.67	0.50	15	0.65	
<i>pgtE</i>	0.50	0.50	17	0.68	TTGA
<i>mgtC</i>	1.00	0.67	16	0.91	TGAC
<i>pipD</i>	0.67	0.50	17	0.72	
<i>pagC</i>	0.83	0.33	17	0.71	

Table S3. Bacterial strains used in this study.

Strain or plasmid	Description	Ref. or source
<i>S. enterica</i> serovar Typhimurium		
14028s	wild-type	[14]
EG13918	<i>phoP</i> -HA	[15]
DH5 α	F ⁻ <i>supE44</i> Δ <i>lacU169</i> (ϕ 80 <i>lacZ</i> Δ M15) <i>hsdR17 recA1 endA1 gyrA96 thi-1 relA1</i>	[16]
Plasmids		
pMS201	pSC101 <i>ori</i> Km ^R , promoterless GFP <i>mut2</i> vector	[17]
pSGF101	pSC101 <i>ori</i> Km ^R , P <i>phoP</i> -GFP <i>mut2</i>	This work
pSGF102	pSC101 <i>ori</i> Km ^R , P <i>pmrD</i> -GFP <i>mut2</i>	This work
pSGF103	pSC101 <i>ori</i> Km ^R , P <i>slyB</i> -GFP <i>mut2</i>	This work
pSGF106	pSC101 <i>ori</i> Km ^R , P <i>ugtL</i> -GFP <i>mut2</i>	This work
pSGF107	pSC101 <i>ori</i> Km ^R , P <i>pagC</i> -GFP <i>mut2</i>	This work
pSGF109	pSC101 <i>ori</i> Km ^R , P <i>pagK</i> -GFP <i>mut2</i>	This work
pSGF111	pSC101 <i>ori</i> Km ^R , P <i>yobG</i> -GFP <i>mut2</i>	This work
pSGF112	pSC101 <i>ori</i> Km ^R , P <i>ugd</i> -GFP <i>mut2</i>	This work

Table S4. Promoter-specific primers used in DNaseI Footprinting and SI nuclease-protection assay

Number	Name	Sequence
6098	<i>A1: mig-14</i>	5' GCGGCATTAAACCGTCGCTTTTCAC 3'
6099	<i>A2: mig-14</i>	5' GCCGTATTGCGTAAACACCTC 3'
9681	<i>A3: mig-14</i>	5' CTGTATCGCTACTGGATGGT 3'
9682	<i>A4: mig-14</i>	5' CCTCCCGGTATAGGGAAAAG 3'
9199	<i>B1: pagK</i>	5' GGATGGGAGCCAGTAAGGA 3'
9200	<i>B2: pagK</i>	5' CGCTCTTAACGTGTTTAATAATAC 3'
9220	<i>C1: yrbL</i>	5' TAAACCTCCCCCTCTATAC 3'
4519	<i>C2: yrbL</i>	5' GTCACGGTTGTAAATGAC 3'
9205	<i>D1: yobG</i>	5' CAGGGCAAACAATACAATTGCGC 3'
9204	<i>D2: yobG</i>	5' CGATGCCGAGAACGACCCATC 3'
5598	<i>E1: pagP</i>	5' AAATCACGGCGCGGTTATTTTTTCATGTC 3'
6527	<i>E2: pagP</i>	5' GAAGGAATAGCCAGCCATGG 3'
6102	<i>F1: slyB</i>	5' GGCCCGATTTTCATACCTCTTCC 3'
6103	<i>F2: slyB</i>	5' ACCCCATCAGTGAAACGGCCAG 3'
9206	<i>G1: pdgL</i>	5' CTGTACTACTGATTATCAAATG 3'
9207	<i>G2: pdgL</i>	5' CAGGCTGAACAGGAGGAAAG 3'
7981	<i>H1: pagD</i>	5' CCCGCTCCGATTTAATCT 3'
7982	<i>H2: pagD</i>	5' CTGGCCAAAACATGGAATG 3'
8201	<i>I1: virK</i>	5' CGCTTGCCGCCTTCCTGCAACT 3'
8202	<i>I2: virK</i>	5' CGATAAGGCGTCTGCGCCATG 3'
9683	<i>I3: virK</i>	5' CTTGCCGCCTTCCTGCAACT 3'
9684	<i>I4: virK</i>	5' CATATCACTTTGCTGCATC 3'
9297	<i>J1: ompX</i>	5' GTCAGTACGAGTAAAGGTGGCA 3'
9298	<i>J2: ompX</i>	5' CCAGTGCTGAAAGACATGCA 3'
8203	<i>K1: pipD</i>	5' CGCTATGCCGATGAACATAATATC 3'
8204	<i>K2: pipD</i>	5' GCCATCTTCGTTGCGCGCGA 3'
8431	<i>L1: ybjX</i>	5' ATCATTCGCCAGCCAGCCATC 3'
8432	<i>L2: ybjX</i>	5' CGAAAGCTGGCTTTACGCCAG 3'
6891	<i>M1: mgtC</i>	5' AATTCATGCAGGAGTAATATG 3'
1250	<i>M2: mgtC</i>	5' GAATTCTGGAAGAATAAGTACGTGC 3'
9732	<i>N1: rstA</i>	5' AGAGGAATTAATCCGGCGA 3'
9733	<i>N2: rstA</i>	5' ACGCGGCTCAACAATGACATC 3'

Table S5. Promoter-specific primers used in SI nuclease-protection assay

Number	Name	Sequence
4518	<i>C3: yrbL</i>	5' GATGACAGTCACCCCAAAACC 3'
4519	<i>C2: yrbL</i>	5' GGTCACGGTTGTAAATGAC 3'
9198	<i>J3: ompX</i>	5' CGGCGGTTGAGGGTTCGTTGA 3'
9202	<i>J4: ompX</i>	5' CCAGAACAGCGGCCAGTGCTG 3'
8203	<i>K1: pipD</i>	5' CCGCTATGCCGATGAACATAATATC 3'
9575	<i>K3: pipD</i>	5' GCGATGGGATGAATAACCTTATG 3'
6005	<i>M3: mgtC</i>	5' GGAATTTATTGTTTAATGATTTTCAGACG 3'
6002	<i>M4: mgtC</i>	5' CGCGTGTTGTAAACATTATTGGGATG 3'

Table S6. Primers used to construct the promoters in GFP reporter plasmids

Number	Name	Sequence
4811	<i>phoP</i>	forward 5' CGGCTCGAG AAGAGTTGACCCGTGGCAAGCGTG 3'
4432	<i>phoP</i>	reverse 5' CGGGATCC TGGCGTAATAATGCATTATCCTC 3'
4842	<i>rstA</i>	forward 5' CGGCTCGAG AAATAAGGAATGATCGACCAC 3'
5231	<i>rstA</i>	reverse 5' CGGGATCC CAGATAAGCGGCAATGAGAG 3'
8160	<i>rstA3</i>	forward 5' CGGCTCGAG CGGAACCGCTCTCGTTTAGA 3'
5231	<i>rstA</i>	reverse 5' CGGGATCC CAGATAAGCGGCAATGAGAG 3'
5228	<i>pagK</i>	forward 5' CGGCTCGAG AACAGCAGAAGAGTTGTTATG 3'
5227	<i>pagK</i>	reverse 5' CGGGATCC GAAGATGGTAATATTTAAACC 3'
4737	<i>mgtC</i>	forward 5' CGGCTCGAG GAGCTATCGCCGGTATTAAGCAGG 3'
4736	<i>mgtC</i>	reverse 5' CGGGATCC CATGAACATGACGATTACCT 3'
8142	<i>phoP4</i>	forward 5' TCGA CTATTTGTCTGGTTTATTAAGTGTTCCTCCCAAAGCACCATAATCAACGCTAGACTG 3'
8143	<i>phoP4</i>	reverse 5' GATC CAGTCTAGCGTTGATTATGGTGCTTTGGGGATAAACAGTTAATAAACAGACAAATAG 3'
8144	<i>rstA4</i>	forward 5' TCGA AACCGCTCTCGTTTAGAAAAGATTTATGGAAGGGAAAACAGAGGCGGTGTATGTTGGCGTTTTCTA 3'
8145	<i>rstA4</i>	reverse 5' GATC TAGAAAACGCCAACATACACCGCCTCTGTTTTCCCTTCATAAATCTTTTCTAAACGAGAGCGGTT 3'
8146	<i>pagK4</i>	forward 5' TCGA TTATTTAAGTTAAATATTTTATAAATGGTTTTTATTTACTCACCTGATGGTAATGAATAACGTTTAATATCTATAGTAAAGG 3'
8147	<i>pagK4</i>	reverse 5' GATC CCTTTACTATAGATATTAACGTTATTCATTACCATCAGGTGAGTAAATAAAAAACATTTATAAAATATTTAACTTAAATAA 3'
8148	<i>mgtC4</i>	forward 5' TCGA ATCAAACAACTTAAACAGAACGTCATAAACCCGCCTTTGCACTTTACGGAACATATTGGCTGACTATAATAAGCGCAAATT 3'
8149	<i>mgtC4</i>	reverse 5' GATC AATTTGCGCTTATTATAGTCAGCCAATATGTTCCGTAAAGTGCAAAGGCGGGTTTAGTGACGTTCTGTTTAAGTTTGTGTTGAT 3'
8150	<i>phoP4-rev</i>	forward 5' TCGA CTATTTGTCATAAACAGTTAATAAACACCCCAAAGCACCATAATCAACGCTAGACTG 3'
8155	<i>phoP4-rev</i>	reverse 5' GATC CAGTCTAGCGTTGATTATGGTGCTTTGGGGTGGTTTATTAAGTGTTCCTGACAAATAG 3'
8162	<i>rstA4-rev</i>	forward 5' TCGA AACCGCTCATAAATCTTTTCTAAACGAGGAAGGGAAAACAGAGGCGGTGTATGTTGGCGTTTTCTA 3'
8165	<i>rstA4-rev</i>	reverse 5' GATC TAGAAAACGCCAACATACACCGCCTCTGTTTTCCCTTCCTCGTTTAGAAAAGATTTATGAGCGGTT 3'

Table S7: Genome-wide analysis of PhoP activated genes in *Klebsiella pneumoniae* using gene expression measured by microarray experiments of wild-type (percentage of gene covered by expressed probe pairs; averaged and maximum values of expressed probe pairs) and phoP mutated strains (Nimblegen tiling arrays), and promoter binding by the PhoP protein, measured by a ChIP-chip assay and identified by using the D&C method [1], where more than one PhoP box can be protected under the same ChIP peak (Position is relative to the upstream end of the corresponding promoter region; C indicates reverse orientation; Score is defined in [0,1]).

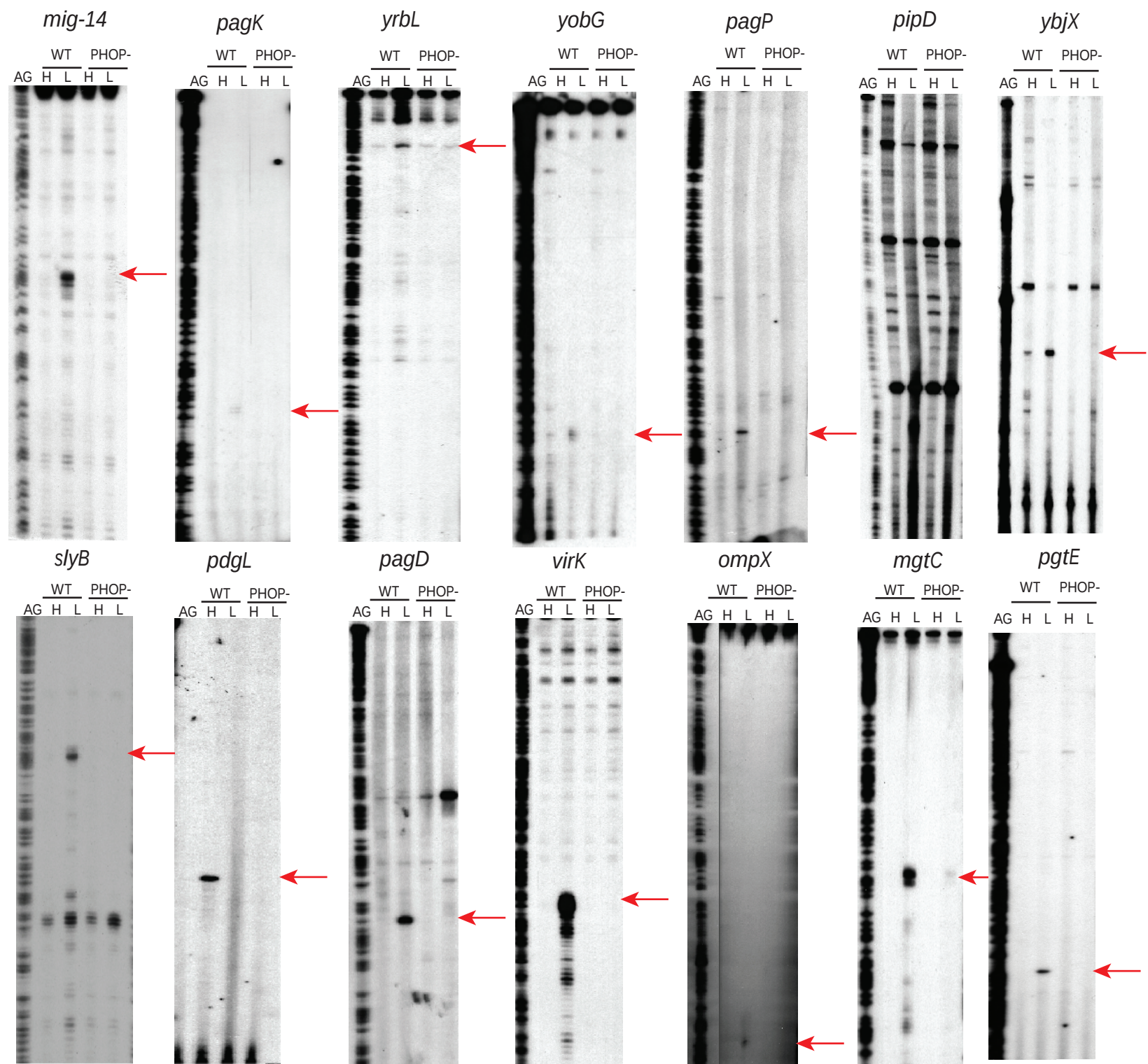
Gene			Expression			ChIP		1st Submotif		2nd Submotif		
Code	Name	Orient.	Coverage %	Average	Maximum	Position	Position	Submotif	Score	Position	Submotif	Score
KPN_04773	N/A	-	100	3.79	5.26	126	209C	3	0.97	182	3	0.7
KPN_03847	<i>yfbE</i>	-	100	10.75	22.35	8	181	5	0.86	79	1	0.85
KPN_02444	<i>pmrD</i>	+	100	5.92	8.61	61	176C	4	0.77			
KPN_01136	<i>phoP</i>	-	100	13.71	29.55	100	209	8	0.72	220	1	0.56
KPN_01979	<i>slyB</i>	+	100	2.43	2.88	100	125	2	0.72	82C	12	0.56
KPN_02997	N/A	-	100	3.24	3.80	53	287C	1	0.65	298C	11	0.56
KPN_00873	<i>ybjG</i>	-	100	5.11	11.01	84	134	7	0.51			
KPN_02355	N/A	+	98	4.19	9.87	22	76C	10	0.76	65C	3	0.75
KPN_01508	N/A	+	96	2.60	3.25	143	125	9	0.59			
KPN_02050	N/A	-	95	5.78	8.59	100	194C	12	0.71			
KPN_01450	N/A	-	92	4.45	10.23	5	138	8	0.7			
KPN_04654	<i>mgtA</i>	+	79	3.12	11.85	312	70	4	0.88			
KPN_02341	<i>yebO</i>	-	63	1.84	5.16	283	93	10	0.75			
KPN_03285	<i>fimB</i>	+	35	0.89	4.16	420	126	6	0.62	513	8	0.52
KPN_00559	N/A	+	26	0.58	2.79	346	9	2	1			
KPN_02051	N/A	+	17	0.44	3.00	223	249	12	0.71			
KPN_01847	<i>sfcA</i>	+	8	0.17	2.17	172	62	11	0.57			
KPN_01521	<i>rstA</i>	-	7	0.16	2.25	93	11C	11	0.83			
KPN_01602	<i>ygbM</i>	-	7	0.17	2.53	16	24	10	0.54			
KPN_01492	N/A	-	4	0.09	2.30	49	160	1	0.66			
KPN_02399	N/A	+	4	0.09	2.48	124	278	6	0.58	60C	5	0.54
KPN_02443	N/A	-	3	0.10	3.13	123	79	4	0.77			
KPN_03124	<i>pyrG</i>	-	3	0.06	2.11	236	20	7	0.5			
KPN_00564	N/A	+	3	0.06	2.15	148	28	5	0.51			
KPN_00843	<i>ompX</i>	+	1	0.01	2.42	252	65	12	0.86	297	5	0.6

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A

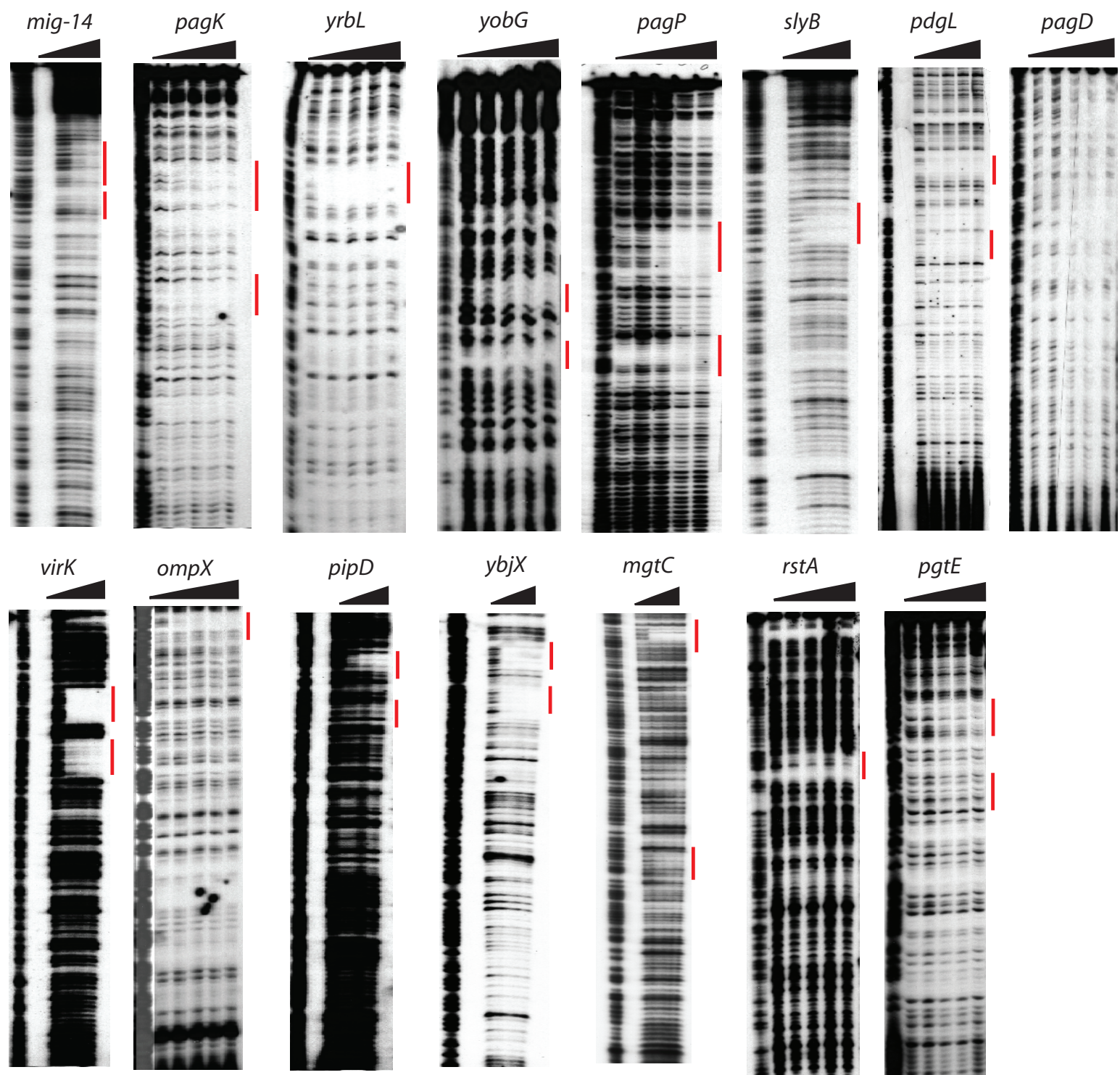
B**Figure S1**



Figure S2

A

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phoP AAGAGTTGACCCGTGGCAAGCGTGTGTATGCCGAAGGAATGAAACAGTTTATTGATAGTCTGGCCCTGCCGAAGCAGAAAAACGCGCCTTAAAGCCATGACGCCGGCAAATTAT
ATCGGTGCGCTGTGACTCTGGTCGACGAACCTTAATAATGCCTGCCTCACCCTCTTTTCTTCAGAAAGAGGGTGACTATTTGTCTggtttattaactgtttaTCCCCAAAGCACC
ATAATCAACGCTAGACTGTTCTTATTGTTAACACAAGGGAGAAGAGATGATGCGCGTACTGGTTGTAGAGGATAATGCATTATTACGCCA

rstA AAATAAGGAATGATCGACCACATGCTAAAGGCGATGGTGGTACGCATCGCGTCGATGCCCCGCTCGCTGGCCACGATATAGTGC GCGATAAGCGCAAACGTTGGAACAGAGGAA
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ybjX4 TCGAGCAAAATTTTAAACGAAAgtaaacggtatctaacaacAACTGACGCAATAAAGATAGCGCCGGCTgtattgacgattggttaatGTTTCCTGTGGTTCAATCTCGACAATCT
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virK4 TCGATTTATTACCGCCATTgataaactgtttaacaacaTCGTCTGTACAGACCTTCTtctgtgcctttacgtttaaCTCAATCAGGCTACCGTCTCGGTTATAAGT
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mgtC GAGCTATCGCCGGTATTAAGCAGGAATTTATTGTTTAAATGATTTTCAGACGAGCTTGTATCGACATAAATATTGTCATTTTTTTGTCATAAGAAAAAATcaaacaaacttaaa
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phoP4/2 TCGATTTATTACCGCCATTgataaactgtttaacaacaTCGTCTGTACAGACCTTCTtggtttattaactgtttatCCCCAAAGCACCATAATCAACGCTAGACTGGATCC

B

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phoP4-rev TCGACTATTTGTCataaacagttaataaaccaCCCCAAAGCACCATAATCAACGCTAGACTGGATCC
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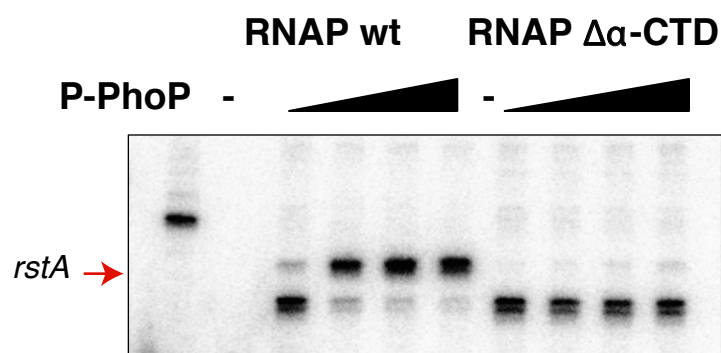
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C

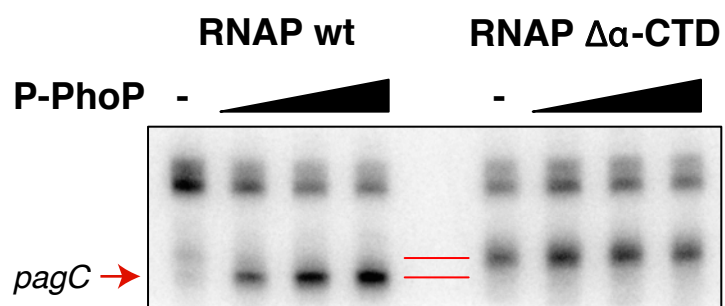
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<i>phoP4+11nt-10</i>	TCGACTATTTGTCtggtttattaactgtttatGGAAGGGAAAACCCCAAAGCACC <u>TATGTT</u> CAACGCT A AGACTGGATCC
<i>phoP4-rev-C</i>	TCGACTATTTGTCataaacagttaataaaccaAACGTCACTAAACCCGCCTTTGCACTT <u>TACGGT</u> TATCCCCAAAGCACCATAATCAACGCT A AGACTGGATCC
<i>phoP4-rev-C-10</i>	TCGACTATTTGTCataaacagttaataaaccaAACGTCACTAAACCCGCCTTTGCACTT <u>TACGGT</u> TATCCCCAAAGCACC <u>TATAAT</u> CAACGCT A AGACTGGATCC
<i>phoP4-rev-K</i>	TCGACTATTTGTCataaacagttaataaaccaTTTTTATTTACTCACCTGATGGTAACCCCAAAGCACCATAATCAACGCT A AGACTGGATCC
<i>phoP4-rev-K-35</i>	TCGACTATTTGTCataaacagttaataaaccaTTTTTATTTACTCT <u>TGACAT</u> TGGTAACCCCAAAGCACC <u>CATAAT</u> CAACGCT A AGACTGGATCC
<i>mgtC4-P-box</i>	TCGAataaacagttaataaaccaAACGTCACTAAACCCGCCTTTGCACTT <u>TACGGA</u> ACATATTGGCTGACT <u>TATAAT</u> AAGCG C AAATTGATCC
<i>phoP4-rev-C</i>	TCGACTATTTGTCataaacagttaataaaccaAACGTCACTAAACCCGCCTTTGCACTT <u>TACGGT</u> TATCCCCAAAGCACC <u>CATAAT</u> CAACGCT A AGACTGGATCC

Figure S3

A



B



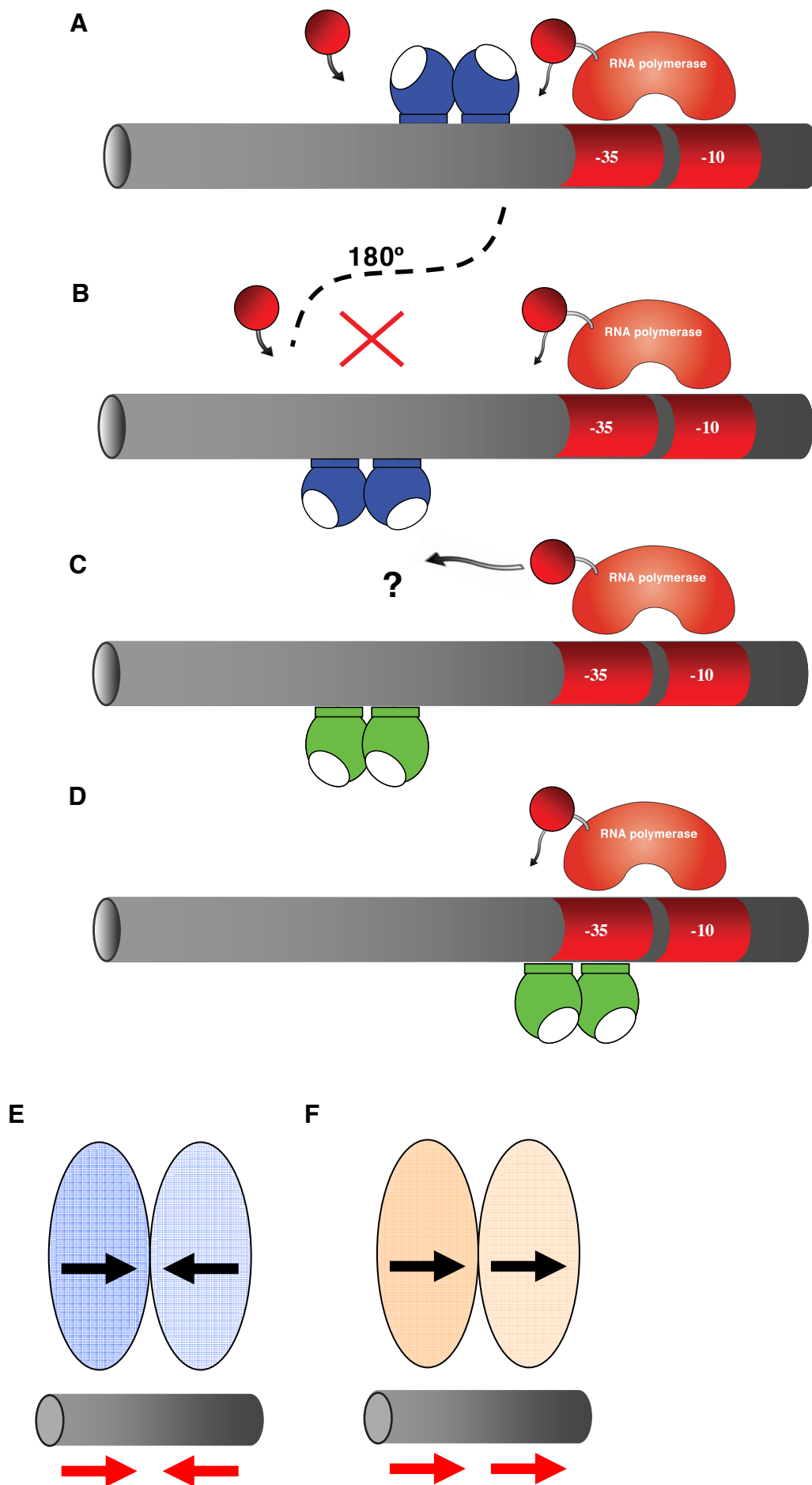


Figure S5

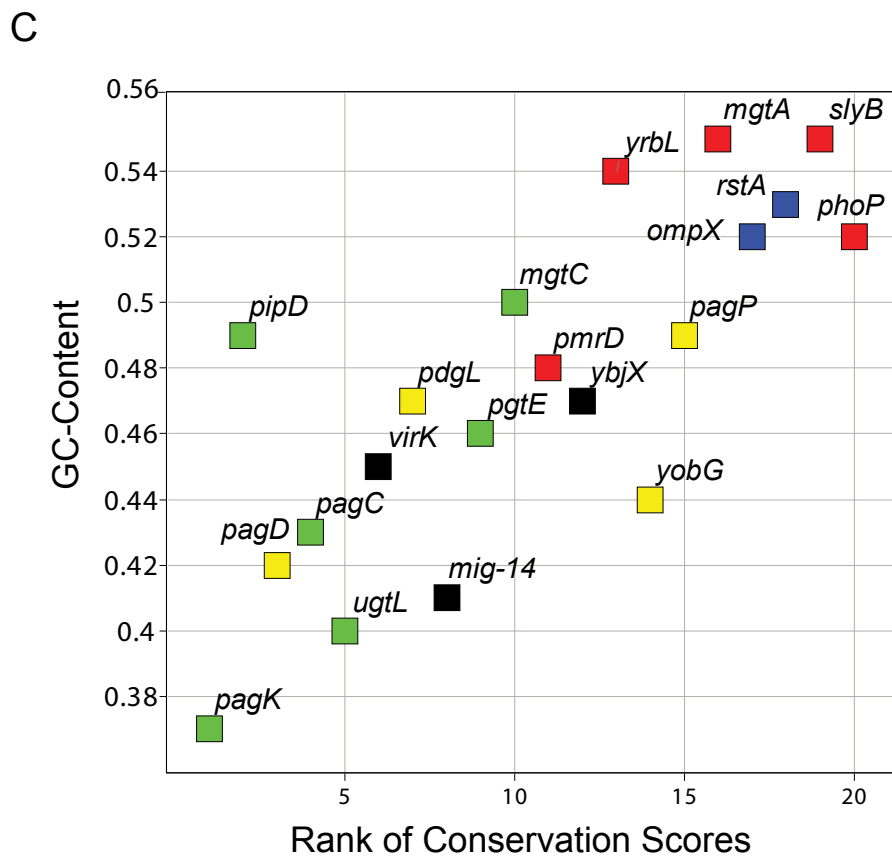
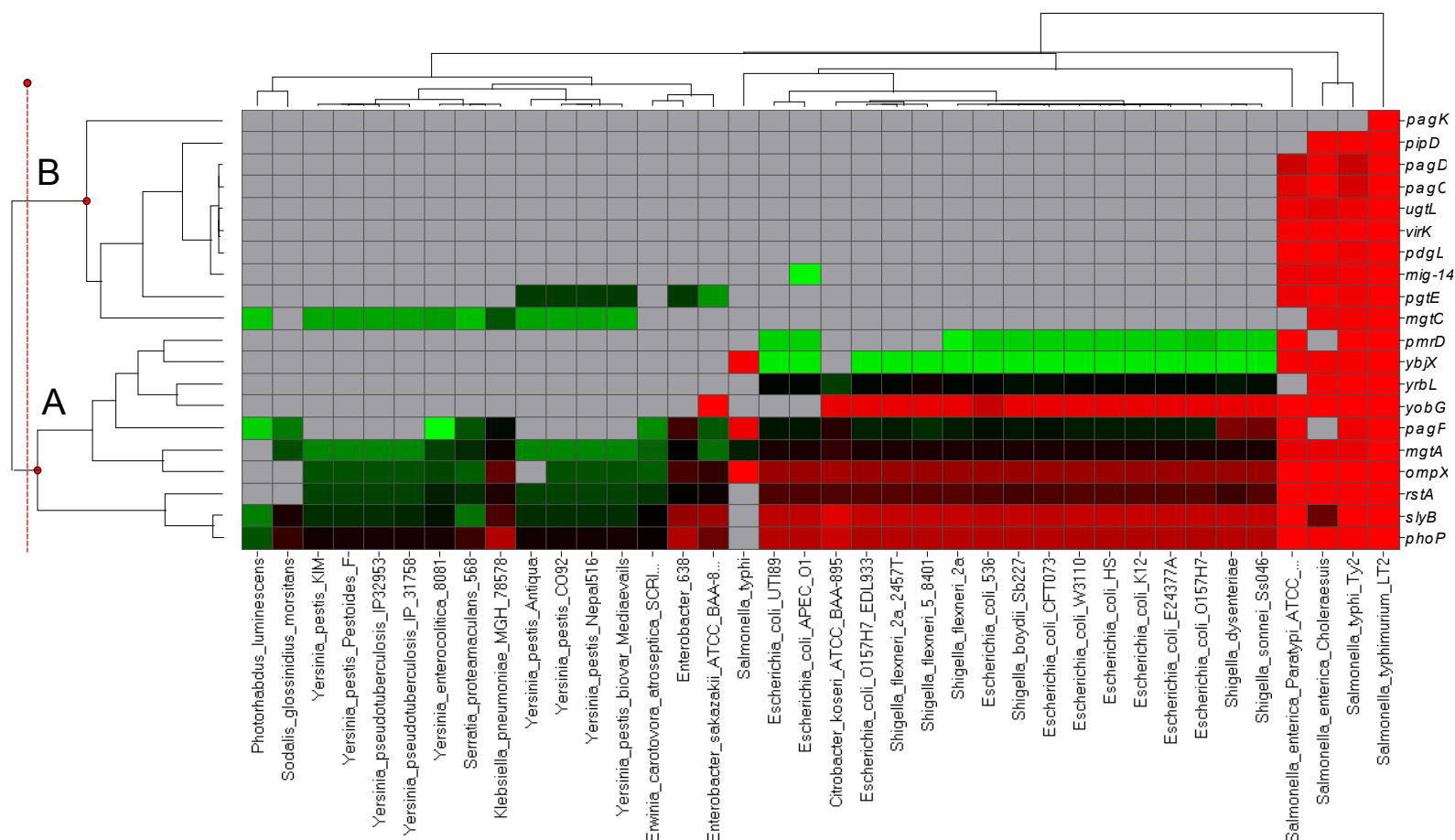


Figure S6